

Figure 1. Alignment between human PMS2 (humPMS134) and Arabidopsis thaliana homologue of PMS2 (AtPMS2) DNA sequences. Similarity is 48.1%; identity is 48.1%. Black boxes show identical nucleotides.

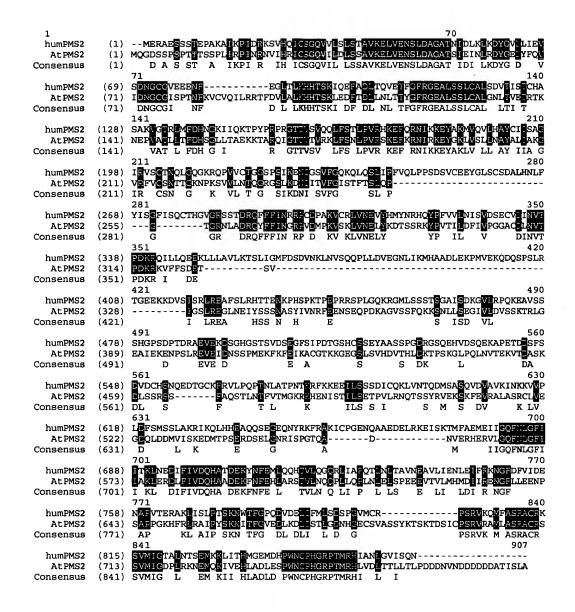


Figure 2. Alignment between human PMS2 (humPMS134) and Arabidopsis thaliana homologue of PMS2 (AtPMS2) amino acid sequences. Similarity is 41.5%; identity is 31.1%. Black boxes show identical residues.

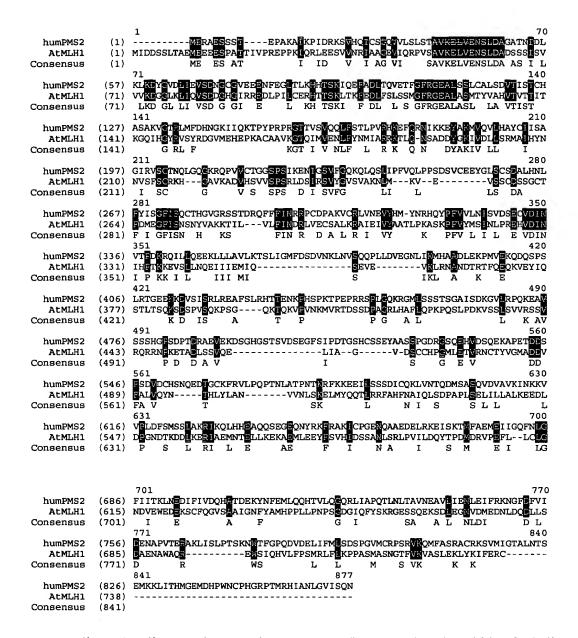


Figure 3. Alignment between human PMS2 (humPMS2) and Arabidopsis thaliana PMS2 homologue MLH1 (AtMLH1) amino acid sequences. Similarity is 30%; identity is 18.4%. Black boxes show identical residues.

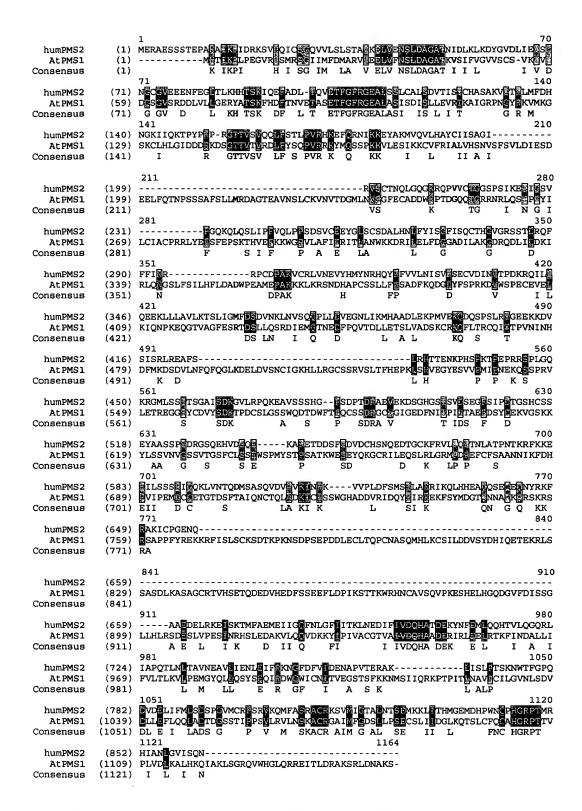


Figure 4. Alignment between human PMS2 (humPMS2) and Arabidopsis thaliana PMS2 homologue PMS1 (AtPMS1) amino acid sequences. Similarity is 24.4%; identity is 15%. Black boxes show identical residues.

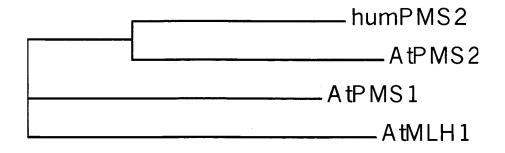


Figure 5. Phylogenetic tree of the Arabidopsis thaliana PMS2 gene homologues.

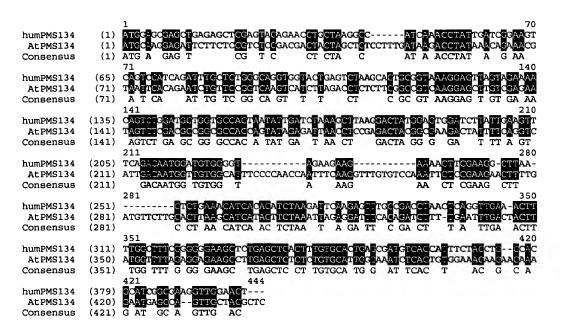


Figure 6. Alignment between human PMS134 (humPMS134) and Arabidopsis thaliana homologue of PMS134 (AtPMS134) DNA sequences. Similarity is 53.2%; identity is 53.2%. Black boxes show identical nucleotides.

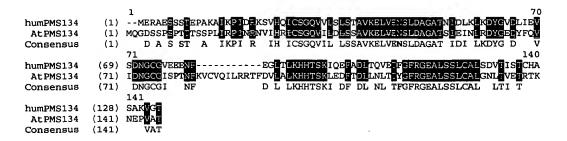


Figure 7. Alignment between human PMS134 (humPMS134) and Arabidopsis thaliana homologue of PMS134 (AtPMS134) amino acid sequences. Similarity is 65.1%; identity is 50.7%. Black boxes show identical residues.

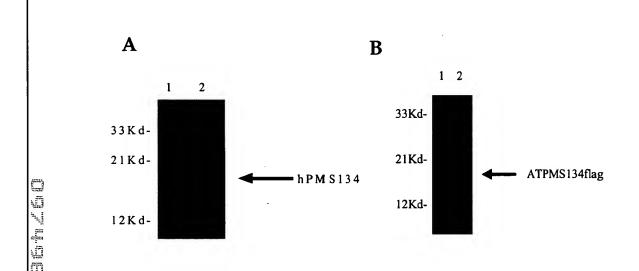


Figure 8: Western blot analysis of bacteria expressing the human PMS134 dominant negative gene (Panel A, lane 2) or the Arabidopsis thaliana dominant negative gene (Panel B, lane 2). Panel A, lysates from bacteria were loaded onto SDS-PAGE gels and probed with an antibody against the human PMS2 N-terminus. Panel B, lysates from bacteria were loaded onto SDS-PAGE gels and probed with an antibody against the flag epitope placed on the C-terminus of the Arabidopsis PMS134 gene. Lane 1 is bacteria

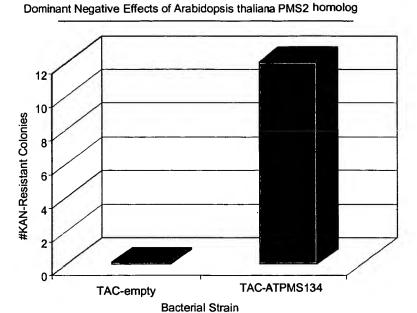


Figure 9. Expression of the *Arabidopsis thaliana PMS134* gene produces hypermutability in bacteria leading to the generation of new phenotypes. Briefly, bacteria containing the empty vector or the TAC ATPMS134 expression vector were grown and plated on kanamycin containing Lbagar plates. The host bacteria are susceptible to KAN bactericidal activity. Bacterial cultures expressing the hPMS134 gene resulted in genetic alteration of the bacterial host and the generation of clones that are KAN resistant.

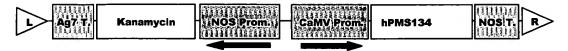


Figure 10. Schematic map of the pCMV-hPMS134-Kan binary plant expression vector. Ag7 T. and NOS T. = gene 7 and Nopaline Synthase poly(A) signals, respectively. NOS Prom and CaMV Prom = Nopaline Synthase and Cauliflower Mosaic Virus promoters, respectively. L and R = left and right T-DNA border repeats, respectively. Arrows indicate direction of transcription.

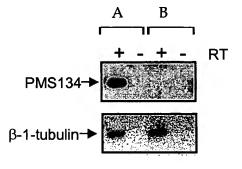


Figure 11. Expression of hPMS134 in Arabidopsis Thailana. Message analysis for T1 plants shows steady state expression of dominant negative MMR genes in PMS134-Kan plants (A) while none is observed in control plants (B). Tubulin was used as an internal control to monitor sample loading and integrity.

A= PMS134 expressing plants B= pBI-121 control plants

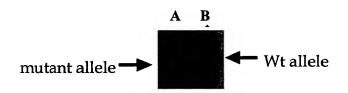


FIGURE 12. Expression of PMS134 expression causes MI in plants. Microsatellite analysis of Nga280 in PMS134 expressing plants (Lane A) found the generation of a mutant allele in contrast to control plants (Lane B).

Figure 13. The plant on the left is a wild type A. thaliana and the one on the right is MMR defective. Seeds from the MMR defective plant have been obtained and offspring have the same "double-meristem" trait.



Normal MMR-